

# Gastric carcinoma in canines and humans, a review

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## Abstract

Gastric carcinoma (GC) is the most common neoplasm in the stomach of dogs. Although incidence in the general population is reported to be low, breed-specific GC has a high incidence. Median age at presentation ranges from 8 to approximately 10 years. The disease is mostly located in the lesser curvature and antropyloric region of the stomach. Unfortunately, diagnosis is usually made when the disease is at an advanced stage and, therefore, prognosis is poor. Due to similarities in clinical presentation, diagnosis, histology and prognosis, canine GC may serve as a valuable model for human GC. Extensive pedigrees of canine gastric carcinoma cases could reveal insights for human gastric carcinoma. Putative species differences include the role of *Helicobacter* in pathogenesis, the wide array of genetic data and screening available for humans, and treatment protocols that are available for human GC.

## Keywords

clinical pathology, comparative oncology, genetics, pathology, small animal

## Introduction

The overall prevalence of gastric carcinoma (GC) in dogs is low although, in a small number of breeds, a higher incidence has been reported.<sup>1</sup> This multi-practice Norwegian study is the only current study comparing the relative in-breed morbidity of gastric cancer. Other case series have variably stated these breed dispositions, or not found any, but may be biased by both regional breed prevalence, small groups, and possible differential referral behaviour.<sup>2–5</sup> Canine GC can be an opportunity as spontaneous model for human GCs. Potential advantages for such comparative research include the fact that dogs share the same environment as humans, the fact that within pedigrees genetic variance is reduced compared to man, and the fact that lifespan is relatively shorter.<sup>6</sup> This article gives an overview of the current knowledge of the clinical and genetic characteristics of GC in dogs and humans. It will address the complete scope of aetiology, clinical presentation, diagnosis, therapy and prognosis and genetics of canine and human GC

(Table 1), but will not be an in-depth review of human GC.

## Aetiology

Human GC ranks as the fifth most prevalent cancer worldwide, and is the third cause of cancer-related death in men and fifth in women.<sup>7</sup> Cancer of the gastro-oesophageal junction (GEJ) is considered a separate group with different epidemiology and aetiology and is outside the scope of this review.<sup>8,9</sup> The aetiology of human GC is complex and incompletely understood. Besides hereditary cancer syndromes, known risk factors include smoking, alcohol consumption, high salt and fat consumption, obesity, low fruit and vegetable consumption, ageing, low economic status, other gastric diseases, and *Helicobacter pylori* infection.<sup>10,11</sup> *H. pylori* infection is the best studied of these and causes a three-to-six-fold increase in gastric cancer risk. About 1–3% of people infected with *H. pylori* develop gastric cancer. It causes chronic

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**Table 1.** Similarities and differences between human and canine gastric carcinoma

Variables	Human	Canine
Incidence	Fourth most prevalent cancer. Especially in Eastern Asia Familial cases: very rare	<1% of all cancer cases Although very high incidence in Dutch Belgian Shepherds.
Age	Middle age to older. Familial cases may be younger	Middle age to older
Sex	Male predilection	Male predilection
Race/breed	Incidence higher in Asians, outside of Asia as well. In addition significant differences in gene expression profiles.	Strong breed predilection in Belgian Shepherds and some other breeds.
Clinical signs	Late in disease	Late in disease Vomiting, anorexia, weight loss
Location	Not including cardia/GEJ tumours; mostly antro-pyloric. In the body and fundus, mostly along the minor and major curvature	Lesser curvature and pylorus. Extension throughout stomach possible
Metastasis	>50% at diagnosis	70–90% at diagnosis
Diagnosis	Mainly by endoscopy	Mainly by endoscopy
Prognosis	Very poor. Western populations: 5-year survival rate 25–30%	Very poor
Histological subtyping	According to Lauren and WHO	According to WHO and Lauren
Treatment	Endoscopic mucosal resection in early gastric cancer Surgical resection in combination with (neo)adjuvant chemotherapy. Prophylactic gastrectomy in CDH1 positive individuals	Surgical resection if possible.
Aetiology	Complex. Important role <i>H. pylori</i> infection. In familial cases not.	Unknown. Suspected complex. Strong genetic component in certain breeds/ pedigrees.
Genetics	Familial cases: HDGC – CDH1. MAP3K, Alpha-E-catenin. Sporadic cases: genetic susceptibility SNPs in MUC1, PSCA and multiple candidate genes including IL17-A, TNF and other interleukins.	No germline mutations found
Screening	Nationwide screening programs in a number of countries, including Japan and Korea. Upper endoscopy and contrast X-ray series.	Not performed at this moment

active inflammation, mucosal damage and altered gene expression and epigenetic changes in multiple genes, eventually leading to carcinogenesis.<sup>12–15</sup>

The intestinal type (IGC) and diffuse type (DGC) of GC are believed to result from distinct pathogenetic pathways. The IGC form develops after stepwise progression from chronic gastritis to atrophic gastritis, metaplasia, dysplasia to intestinal type carcinoma. DGC, however, is thought to arise *de novo* by downregulation of *CDH1*.<sup>11,16</sup> Early stages of DGC are characterised by, often multiple, foci of superficial signet ring carcinoma *in situ*.<sup>16–18</sup> Multiple hereditary cancer syndromes increase risk of GC. Hereditary diffuse gastric cancer (HDGC) carries a cumulative lifetime risk of up to 70% in men and 56% in women of developing GC for mutation carriers.<sup>19,20</sup> *H. pylori* infection does not

appear to be associated with HDGC.<sup>21</sup> Other, rare, hereditary cancer syndromes increasing the risk of GC are: Lynch syndrome, gastric adenocarcinoma and proximal polyposis of the stomach (GAPPS), familial adenomatous polyposis, Li-Fraumeni syndrome and Peutz-Jeghers syndrome.<sup>22–24</sup>

Similar to human GC, the aetiology of canine GC is complex and poorly understood, but it is believed that both environmental and genetic factors play a role.<sup>1,25</sup> Long-term administration of N-Nitrosamines can lead to induction of GC, although the relevance of this in clinical settings is unclear.<sup>26</sup> The strong breed predisposition and familial occurrence described for several canine breeds, including the Belgian Shepherd dog, indicate the importance of genetic factors in the aetiology of gastric cancer.<sup>1,25,27</sup> In addition, some

very small subgroups of canine gastric cancer may have different aetiologies. For example few cases of hypertrophic gastritis (Ménétrières-like disease) progressing into superficial signet-ring-type GC are reported.<sup>28,29</sup> In the Norwegian Lundehunde, atrophic gastritis, characterised by reduction in parietal cells and hyperplasia of neuroendocrine cells, is reported to be associated with gastric tumour development. In these dogs, hypergastrinaemia secondary to atrophy may be important in carcinogenesis.<sup>30,31</sup> Unlike in humans, precursor lesions are not frequently described in dogs that develop GC and no convincing relationship could be demonstrated between gastric polyps that are sporadically identified, *Helicobacter*, and GC.<sup>32</sup> *H. pylori* is infrequently found in the stomach of dogs, likely due to the different glycosylation profile and the ability of different strains to adhere to specific host glycan receptors.<sup>33</sup> However, different 'Non-*Helicobacter-pylori*-type *Helicobacter*' species (NHPH) are frequently encountered in the stomach of both healthy dogs and those showing signs of gastrointestinal disease.<sup>34,35</sup> The pathogenic significance of these *Helicobacter* species remains controversial in dogs, since a clear relationship between NHPH and gastritis has not been established.<sup>36</sup> Therefore, a possible role in the aetiology of GC is questionable.

## Clinical presentation of GC

### Humans

Incidence of GC is particularly high in Eastern Asia with Central and Eastern Europe and South America second and third. Stomach cancer rates are approximately twice as high in males as in females.<sup>37</sup> The vast majority of GCs arise sporadically. Familial clustering is apparent in less than 15% of cases and most of these are not associated with a definitive germline mutation,<sup>38</sup> and hereditary cancer syndromes are associated with less than 3% of gastric cancer cases.<sup>39,40</sup>

The epidemiology and predominant location of GC varies geographically due to a complex of genetic and environmental factors.<sup>11,41</sup> Carcinomas in the body or the corpus of the stomach are typically located along the greater or lesser curvature.<sup>42</sup> Similarly, the location of the tumour in hereditary

forms of GC also appears to vary with racial and geographical differences.<sup>21</sup> GC patients are typically middle-aged to older with most patients aged between 50 and 70 years at presentation. Patients younger than 30 years of age are rarely seen.<sup>43</sup>

Clinical signs of GC in humans tend to emerge late in the development of the disease. Therefore, the cancer is often at an advanced stage at presentation<sup>44</sup> and more than half the patients have lymph node metastasis at first clinical presentation or surgery.<sup>45</sup>

### Dogs

According to a recent Norwegian survey, canine GC accounts for only 0.16% of all reported canine cancer cases.<sup>1</sup> This is in accordance with earlier reports in which GC cases accounted for less than 1% of all canine tumours.<sup>46,47</sup> However, GC is the most common neoplasm in the stomach of dogs compared to the less frequently seen leiomyo(sarco)ma, gastro-intestinal stromal tumours (GIST) and lymphoma.<sup>4,48–51</sup> An increased prevalence of GC is present in several dog breeds with breed predispositions reported for the Staffordshire bull terrier, Rough Collie,<sup>52</sup> Bouvier des Flandres, Standard poodle, Norwegian elkhound,<sup>1</sup> Hovawart and Chow-Chow<sup>4,50</sup> and, most convincingly, in the Belgian shepherd, Tervueren and Groenendael.<sup>1,25,27,53</sup>

GC is typically a disease of middle-aged to older dogs. Median age at presentation ranges from 8 to approximately 10 years,<sup>2,4,52,54–56</sup> although there are anecdotal reports of cases less than 5 years of age.<sup>55</sup> Comparable to humans, GC seems to occur more frequently in male than in female dogs. Seim-Wikse *et al.* reported a statistically different odds ratio (OR) for males versus females of 2.3, with only a male to female ratio of 1.2:1.<sup>1</sup> However, there are other ratios reported varying from 1.4:1 to 2.75:1.<sup>4,25,50,56</sup> This variation might be explained by the use of relatively small numbers of dogs in these studies as well as phenotypical diversity. There are indications that the male:female ratio is different in different subtypes of gastric adenocarcinoma.<sup>47</sup>

As in humans, clinical signs of gastric cancer in dogs are usually mild to absent during the early stages of the disease. The most common

clinical signs are vomiting (40–95% of cases), anorexia (48–52%) weight loss (23–52%) and lethargy (25–28%).<sup>2,4,50</sup> Gualtieri *et al.* found no strong relationship between the extent of gastric wall involvement of the tumour and the severity of vomiting. Haematological and biochemical changes are typically absent or small and may consist of mild anaemia, increased liver enzymes, hypoglycaemia, hyperproteinaemia, hypalbuminaemia and prolonged clotting times.<sup>4,50</sup>

Metastasis has classically been reported to occur at a high rate, and has typically reached 70–90% by the time of diagnosis.<sup>2,4,47,52</sup> A possible explanation for this could be that the early stage of the disease is often missed due to the mild nature of the clinical signs meaning that veterinary advice is not sought. The most common site of metastasis is the regional lymph nodes. More distant sites frequently reported are the liver, lung, duodenum, adrenal glands, pancreas, omentum and spleen. In a few more recent smaller studies, metastasis percentage during presentation was low.<sup>50,57–59</sup> It is unclear whether this is due to lesser malignancy in these cases, earlier presentation or the diagnostic protocol used.

The primary tumour in canine GC is most often located in the lesser curvature and pyloric region of the stomach but can extend throughout the stomach.<sup>2,4,25,47,59</sup> The gastric wall is often markedly thickened and contains abundant fibrous tissue. This scirrhous aspect of the tumour can make the stomach wall non-distensible, a condition sometimes referred to as *linitis plastica*. Ulceration of the mucosal surface is present in about half of the cases and may be characterized by a necrotic centre with a raised rim.<sup>4,47,52</sup>

## Diagnosis

In humans, different diagnostic guidelines vary with the geographic location. However, when GC is suspected, endoscopy with multiple biopsies is recommended consistently in all national and international guidelines.<sup>60,61</sup> Over 90% of tumour cases are diagnosed using endoscopy,<sup>62</sup> partially in conjunction with methods to improve visualization of the mucosa in endoscopy such as confocal endomicroscopy or narrow band imaging, to detect vascular patterns, to improve sensitivity and

specificity.<sup>63,64</sup> There is broad consensus that staging is mandatory for decision making: this can be performed by physical examination, clinicopathological testing (e.g. complete blood count, tests of liver and kidney function), and advanced imaging such as abdominal/pelvic CT (possibly combined with PET-CT) and either CT or radiography of the thorax. However, throughout Europe, endoscopic ultrasonography is the method of choice for the detection of locoregional metastasis and to assess tumour infiltration depth. Laparoscopy, with or without peritoneal wash to detect malignant cells cytologically, is recommended to exclude occult metastasis in surgical candidates.<sup>61,65,66</sup>

In dogs, the clinical signs, signalment and family history are often sufficient to include GC as a differential diagnosis. Furthermore, in breeds with a strong breed predisposition and a large database of confirmed cases, family history may contribute to the suspicion of GC. Abdominal radiographs may show thickening of the gastric wall, loss of (cranial) abdominal detail, cranial abdominal mass effect or no specific findings.<sup>4,50</sup> However, this type of diagnostic investigation is now less commonly performed because of the advent of ultrasonography and CT.

Abdominal ultrasonography enables the morphological assessment of the stomach wall, abdominal lymph nodes, and other possible metastatic sites. This may enable a tentative diagnosis of GC to be made and can direct further diagnostic work-up. The majority of GCs are ultrasonographically sessile masses involving all layers of the stomach, and may extend through the serosal surface. Evidence of ulceration and lymphadenopathy may also be apparent, but ultrasonography has relatively poor sensitivity for detection of such lesions. Further, dogs with GC share many ultrasonographic features with dogs that have gastric lymphoma.<sup>49,67</sup> Although ultrasonography is a valid tool in the diagnostic approach of GC, recent studies found only 50–58% compatibility of ultrasonography in clinical cases of stomach cancer.<sup>50,68</sup>

CT scans are very helpful in assessing stomach wall, lymph nodes, and other indications of metastasis,<sup>50,58</sup> but challenges in interpreting the morphology of the stomach wall persist, most commonly due to gas artefacts or incomplete

distension. This can be overcome by using Helical Hydro CT, which offers enhanced contrast and stomach extension; indeed, in a recent feasibility study, imaging results corresponded well with histology of gastric and lymph node biopsies.<sup>59</sup>

Endoscopy is the diagnostic method of choice for GC,<sup>4,48,69</sup> since the epithelial origin of the lesions typically makes for a high rate of diagnostic accuracy.<sup>53</sup> However, in some cases it can be difficult to differentiate changes associated with GC from inflammation.<sup>70</sup> Diagnostic methods to improve the evaluation of gastric mucosa in endoscopy are not routinely used, but may be available in the future to aid *in vivo* diagnosis of (early) gastrointestinal disease.<sup>71</sup>

Exploratory coeliotomy is the gold standard for diagnosing GC, but is highly invasive. Its main benefits are the fact that it enables full thickness gastric biopsies to be acquired, and also a complete evaluation of abdominal metastasis. A further advantage of coeliotomy is the potential that simultaneous surgical removal can also be considered if indicated.

### Histology

Ninety percentage of human gastric cancers are adenocarcinomas, and these are subdivided by histological appearance using a scheme referred to as the Laurén classification into diffuse and intestinal types<sup>72</sup> or classified according to the WHO-guidelines according to the most dominant histological pattern. The WHO classification recognises four main types: (1) Tubular adenocarcinomas with prominent dilated or slit-like and branching tubules. Tumour cell morphology is columnar, cuboidal, or flattened by intraluminal mucin; (2) papillary adenocarcinomas, well-differentiated exophytic tumours with elongated processes lined by cylindrical or cuboidal cells and a fibrovascular connective tissue core; (3) mucinous adenocarcinomas in which extracellular mucin pools constitute over 50% of the tumour. The tumour cells can form glands of columnar mucous-secreting epithelium or irregular cell clusters. Occasional scattered signet ring cells may be present but do not dominate; and (4) signet-ring cell carcinomas where >50% of the tumour consists of malignant cells containing intracytoplasmic mucin and the nuclei may be

displaced against cell membranes.<sup>42</sup> Histological subtyping is important because of differences in likely pathogenesis, diagnostic testing, therapy and prognosis.<sup>18,61,73–75</sup> The term ‘early gastric cancer’ (ECG) is used for a carcinoma limited to the mucosa or the mucosa and submucosa. In addition, ‘precursor lesions’ may be recognised in asymptomatic patients and in tissue surrounding cancerous lesions. Chronic atrophic gastritis and intestinal metaplasia and dysplasia commonly precede and/or accompany GC of the intestinal type, although only a small portion of these patients progress to develop gastric cancer.<sup>42,76</sup>

In dogs, surgical or endoscopic biopsies are mostly assessed according to a scheme adapted from the human WHO classification with major types based on predominant histological growth pattern: Papillary, tubular, (tubulopapillary,) mucinous, signet ring or undifferentiated. The other classification system commonly used is the ‘Laurén’ classification with the intestinal type tumours forming poorly- to well-differentiated glands and the diffuse type of GC being composed of poorly cohesive cells and lacking glandular structures. The diffuse type tumour will often contain cells with large amounts of intracytoplasmic mucin called signet ring cells.<sup>72,77</sup> Classification according to the Laurén scheme, with less subgroups gives more power to often small case sets and allows for easier translation to human medicine. The clinical and prognostic significance of the subtypes, irrespective of Laurén or WHO-classification systems, in canine GC has not been clearly demonstrated so far.<sup>56</sup>

### Biomarkers

In human medicine, the search for prognostic or predictive biomarkers for GC is important in the field of personalised medicine. Indeed, a number of candidate biomarkers have been discovered. HER-2 is a proto-oncogene encoded by the ERBB2 gene, and is a tyrosine kinase receptor belonging to the epidermal growth factor receptor (EGFR) family.<sup>78</sup> It is an important predictive biomarker for GC.<sup>79</sup> HER-2 testing is recommended in American, European and Asian gastric cancer guidelines in patients with metastatic or recurrent gastric cancer under consideration for palliative chemotherapy. When

score of immunohistochemical staining for HER-2 is 3+, targeted therapy is indicated.<sup>60,66,80–82</sup> The *c-Met* proto-oncogene encodes the *c-Met* receptor. High MET gene amplification and expression in gastric cancer serve as an independent negative prognostic factor. C-Met has also been identified as a possible therapeutic target. A third potential biomarker with possible predictive value is the ligand of PD-1, an immune-inhibitory receptor of the CD28 family which plays a role in tumour immune escape.<sup>79</sup>

The tyrosine kinases HER-2 and EGFR (Her-1) have recently been investigated in canine gastric adenoma and carcinoma. HER-2 positivity (3+ staining) was present in 11 of 19 gastric tumours, independent of histological subtype or malignancy. EGFR expression was present in 42% of samples and significantly more in intestinal-type than those in diffuse-type tumours.<sup>83</sup> The potential implication of these proteins as predictive or prognostic biomarkers needs further investigation and confirmation with gene expression studies. HER-2 immunohistochemistry staining score was found to be poorly correlated with mRNA expression levels in canine mammary tumours, likely due to lack of specificity of the human antibody on canine tissue, or post-translational effects.<sup>84</sup>

Recently, serum gastrin expression was investigated as a possible biomarker in dogs with GC. This was prompted by a case report of high gastrin in a cattedog with hypergastrinaemia and mucinous GC, and the confirmed association between atrophic gastritis and gastric neoplasia in Norwegian Lundehunde.<sup>31</sup> However, serum gastrin was not useful as a single biomarker.<sup>55</sup> In the search for a biomarker to aid diagnosis of GC in dogs, HER-3 expression and CDX-2 expression have also been investigated. The transmembrane glycoprotein HER-3 is overexpressed in various human cancers including stomach cancer.<sup>85,86</sup> The nuclear transcription factor CDX-2 is a reliable immunohistochemical marker for human gastrointestinal adenocarcinoma and metastases.<sup>87,88</sup> Marked HER-3 expression and CDX-2 expression was demonstrated in the canine gastric adenocarcinomas and the lymph node metastasis, and both HER-3 expression and CDX-2 expression was not detected in normal canine gastrointestinal

tissues.<sup>57</sup> Thus, complementing standard histological examination of tissue samples with immunohistochemistry may improve diagnostic accuracy and better identify metastasis.<sup>57,58</sup>

### Therapy and prognosis

In humans, treatment of gastric cancer is highly dependent on clinical staging and varies in invasiveness according to national and international guidelines. Early gastric cancer lesions can be treated by endoscopic resection and have a favourable prognosis. Indeed, national screening programmes in Japan and Korea have identified many more patients with early gastric cancer in these countries.<sup>60,89</sup> Screening may encompass double-contrast radiography studies followed by upper endoscopy or only upper endoscopy.<sup>90</sup>

For all patients with advanced gastric cancer without distant metastasis, curative surgery is the aim. Margins of >5 cm for diffuse gastric cancer and 2–3 cm for intestinal type cancer should be maintained and depending on the grade of cancer D1 lymph nodes (perigastric nodes directly attached along the lesser curvature and greater curvatures of the stomach) or D2 lymph node (D1 plus nodes along the left gastric artery, common hepatic artery, celiac trunk, splenic hilus, and splenic artery) resection is performed.<sup>91,92</sup>

In people carrying a mutation for HDGC, prophylactic total gastrectomy is offered in early adulthood, typically at an age 5 years younger than the youngest affected family member.<sup>75,93</sup>

Peri-operative chemotherapy has been widely adopted as standard care throughout the world. Although in North America chemoradiotherapy is often the first choice, where 5-FU and Leucovorin are combined with radiotherapy.<sup>80</sup> Much used protocols include ECF (epirubicin, cisplatin and 5-FU) or ECX (epirubicin, cisplatin and capecitabine) in Europe and the UK<sup>66,94</sup> and S-1 sometimes with addition of cisplatin or capecitabine plus oxaliplatin in Asia.<sup>60,95</sup> For non-resectable disease, palliative chemotherapy is used, with the addition of trastuzumab in HER2 positive patients.<sup>66,80,82</sup>

The only potentially curative treatment option for GC in the dog is surgical resection. Malignancies should be resected with wide margins of

1–2 cm of apparently normal tissue around the tumour. Unfortunately, in many cases, the extent and location (especially when the lesser curvature is involved) precludes such dissection. Tumours in the antropyloric region may be resected by partial gastrectomy and gastroduodenostomy.<sup>4</sup> Pylorotomy with gastroduodenostomy has a poor long-term survival time in dogs with malignant neoplasia. Median overall survival time of dogs with malignant neoplasia and metastasis was only 33 days (95% CI 14–578).<sup>70</sup> Surgical resection in 10 cases of gastric adenocarcinoma resulted in a median survival time of 72 days (range 3 months to 4 years).<sup>4</sup> Survival after surgical resection alone in eight dogs resulted in a short survival of between 2 days and 10 months.<sup>2</sup> Chemotherapy alone or adjuvant chemotherapy after surgical resection has been described in small case studies and case reports only and may be of limited additional value in advanced disease. Chemotherapy protocols used include carboplatin, doxorubicin, doxorubicin combined with cyclophosphamide and 5-fluorouracil, followed by cisplatin and 5-fluorouracil in combination with cyclophosphamide.<sup>2,50,58,96</sup> In a report of three cases, survival time was 9 weeks, 9 weeks and 7.5 months, respectively.<sup>2</sup> In other reports of a single case, survival time after surgery and adjuvant chemotherapy was 81 and in 114 days, respectively.<sup>50,96</sup>

Total gastrectomy has been performed sporadically in dogs with GC, but ethical questions remain in performing this extensive surgery in dogs with such advanced disease, because of post-operative quality of life and expected prognosis with metastatic disease.<sup>4,97</sup>

Surgery with adjuvant chemotherapy of gastric adenocarcinoma confined to the mucosa or submucosa may have a more favourable prognosis.<sup>58</sup> Poor prognosis of dogs with GC is likely influenced by late presentation. Recent evaluation of more than 80 biopsies at the Department of Pathobiology, Faculty of Veterinary Medicine, Utrecht University, has shown that it is not always possible to determine the extent of neoplastic infiltration into the muscular layers based on the scant tissue collected during endoscopic biopsy (unpublished observations). This additional uncertainty further complicates the decision as to whether or not surgery is indicated.

Furthermore, the less well-organised diffuse-type or signet ring-type tumours are considered to have a less favourable prognosis than the more glandular tumours. There are indications that these tumours might metastasise earlier.<sup>98</sup> To the authors knowledge there are currently no screening programmes in dogs.

## Genetics

### Humans

In humans, most cases of gastric cancer are sporadic without evidence of germline mutations, and arise as a result of the complex interplay between environmental risk factors and (acquired) genetic factors. These genetic factors may be the result of somatic gene mutations, single nucleotide polymorphisms (SNPs) in known candidate genes, chromosomal and microsatellite instability, or changes in miRNA profile or epigenetic landscape. Through numerous studies, a multitude of genes and genomic regions have been implicated in gastric cancer and disease progression. For example, APC, TP53, NME1 have been implicated with loss of heterozygosity.<sup>99</sup> Microsatellite instability is found in up to 38% of sporadic gastric cancer cases and high microsatellite instability is associated with better prognosis.<sup>100,101</sup> SNPs in inflammatory genes have also been extensively studied, particularly in genes involved in the host response to *H. pylori* infection.<sup>102,103</sup> A positive association between IL1 markers and increased risk of gastric cancer have also been confirmed by several meta-analyses.<sup>104,105</sup> However, ethnical and geographical factors, histological subtype (diffuse versus intestinal), and anatomical location of the tumour are also of importance in this association.<sup>39,106</sup> Another gene with an established gene polymorphism increasing gastric cancer risk is IL17A.<sup>107,108</sup> The host response to *H. pylori* infection and its influence on gastric cancer development is also reflected in the MUC1 polymorphism, with the G allele conveying an almost 30% reduced risk of gastric cancer.<sup>109</sup>

The PSCA gene, encoding prostate stem cell antigen identified with a Genome Wide Association Study (GWAS) was shown in multiple patient cohorts to hold a polymorphic genetic variation increasing the risk of gastric cancer

development. PSCA was revealed to have a role in the inhibition of epithelial cell proliferation.<sup>110,111</sup> A high level of evidence for association with risk of developing sporadic GC was confirmed for polymorphisms of PSCA and MUC1, in addition to MTX1, PRKAA1, PLCE1, TGFBR1, PKLR, GSTP1, CASP8 and TNE.<sup>112</sup> In addition to these and other risk-conferring genetic factors, a large number of nonsynonymous somatic mutations are present in gastric adenocarcinomas. Frequently mutated genes include TP53, genes of cell adhesion pathways, chromatin remodelling genes and, in diffuse-type carcinomas specifically, RHOA – a RAS homology family member.<sup>113,114</sup>

HDGC is an autosomal-dominant condition classically caused by a germline mutation in the CDH1 gene. HDGC is a clinical diagnosis based on the criteria of the International Gastric Cancer Linkage Consortium. Established 2015 criteria for CDH1 testing are: (1) Two GC cases, at least one confirmed DGC, (2) One case of DGC under the age of 40, (3) Personal or family history of DGC and lobular breast cancer, one diagnosed under the age of 50.<sup>93</sup> The CDH1 gene encodes E-cadherin an important cell adhesion molecule. The mutation is most frequently heterozygous and can take many forms and involve both coding and non-coding regions.<sup>20</sup> Loss of function of the remaining allele can be caused by genetic and epigenetic changes such as promoter hypermethylation and loss of heterozygosity.<sup>115</sup> CDH1 mutations explain about 10–40% of HDGC cases, with the higher percentages at risk of ascertainment bias.<sup>21,93,116</sup>

A truncating allele in the related protein alpha-E Catenin was identified in a Dutch family with HDGC without CDH1 mutation through exome sequencing.<sup>117</sup> Alpha-E-catenin acts together with  $\beta$ -catenin in connecting the cytoplasmic domain of E-cadherin to the cytoskeleton. Mutations in the catenin gene family, however, did not appear to be of more widespread importance in CDH1-negative HDGC families.<sup>118</sup>

Recently, novel germline mutations in non-CDH1 hereditary gastric cancer have been identified. Multiple genetic mutations in the mitogen-activated protein kinase kinase kinase (MAP3K6) were described in patients with familial gastric cancer from different families.<sup>119</sup> Also

candidate mutations in BRCA2, STK11, SDHB, PRSS1, ATM, MSR1 and PALB2 were identified in individual CDH1-negative HDGC family members.<sup>20</sup> However, in the majority of families with hereditary gastric cancer, the underlying genetic cause remains unknown.

## Dogs

Depending on the patient population at a given research institute, most cases of canine GC are likely to be spontaneous or familial. For instance in the Netherlands, sporadic cases are rare, and the majority of cases occur in interrelated dogs (unpublished observation). Sporadic cases of GC arise through acquired somatic mutations during life, possibly in conjunction with environmental risk factors and epigenetic events. These mutations likely include cell cycle proteins.<sup>56</sup> *KRAS* exon 2 status has been evaluated in canine gastric adenomas and adenocarcinomas in one recent study. The *KRAS* gene encodes a small protein involved in signal transduction between transmembrane receptors (like the EGF-receptor superfamily) and the nucleus<sup>120</sup> and was mutated in one of 14 canine GC cases.<sup>83</sup>

In familial canine GC, germline mutations are thought to be passed on through generations with environmental and epigenetic factors contributing to disease expression. However, to date no known mutations in dogs with GC have been published.

## Conclusion

Similar to human GC, canine GC is a heterogeneous disease with limited treatment options and a poor prognosis. While it is appreciated that genetic and environmental factors play an important role in cases of GC, much remains unknown about the aetiology. Given the frequent occurrence of familial, presumed hereditary, cases of canine GC in some regions, an opportunity exists for research into the genetics of GC. Extensive pedigrees of canine GC cases may provide a valuable model for human hereditary GC. Also due to the unique characteristics of inbred dog populations, a far smaller number of animals and DNA markers are needed.<sup>121</sup> For a better prognosis of canine GC in the future and to take advantage of the vast amount of research conducted in humans, it is important to identify cases



as early as possible in the course of the disease. A possible benefit for cost-effective screening patients with increased risk of canine familial gastric cancer remains to be investigated.

## References

1. Seim-Wikse T, Jorundsson E, Nodtvedt A, Grotmol T, Bjornvad CR, Kristensen AT, *et al.* Breed predisposition to canine gastric carcinoma – a study based on the Norwegian canine cancer register. *Acta Veterinaria Scandinavica* 2013; **55**: 25.
2. Swann HM and Holt DE. Canine gastric adenocarcinoma and leiomyosarcoma: a retrospective study of 21 cases (1986–1999) and literature review. *Journal of the American Animal Hospital Association* 2002; **38**: 157–164.
3. Bilek A and Hirt R. Breed-associated increased occurrence of gastric carcinoma in chow-chows. *Wiener Tierärztliche Monatsschrift* 2007; **94**: 71–79.
4. Gualtieri M, Monzeglio MG and Scanziani E. Gastric neoplasia. *The Veterinary clinics of North America: Small Animal Practice* 1999; **29**: 415–440.
5. Penninck DG. Characterization of gastrointestinal tumors. *The Veterinary clinics of North America: Small Animal Practice* 1998; **28**: 777–797.
6. Rowell JL, McCarthy DO and Alvarez CE. Dog models of naturally occurring cancer. *Trends in Molecular Medicine* 2011; **17**: 380–388.
7. Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J and Jemal A. Global cancer statistics, 2012. *CA: A Cancer Journal for Clinicians* 2015; **65**: 87–108.
8. Li-Chang HH, Kasaian K, Ng Y, Lum A, Kong E, Lim H, *et al.* Retrospective review using targeted deep sequencing reveals mutational differences between gastroesophageal junction and gastric carcinomas. *BMC Cancer* 2015; **15**: 32.
9. Wu H, Rusiecki JA, Zhu K, Potter J and Devesa SS. Stomach carcinoma incidence patterns in the United States by histologic type and anatomic site. *Cancer Epidemiology, Biomarkers & Prevention* 2009; **18**: 1945–1952.
10. Gonzalez CA, Lujan-Barroso L, Bueno-de-Mesquita HB, Jenab M, Duell EJ, Agudo A, *et al.* Fruit and vegetable intake and the risk of gastric adenocarcinoma: a reanalysis of the European Prospective Investigation into Cancer and Nutrition (EPIC-EURGAST) study after a longer follow-up. *International Journal of Cancer* 2012; **131**: 2910–2919.
11. Wadhwa R, Song S, Lee JS, Yao Y, Wei Q and Ajani JA. Gastric cancer-molecular and clinical dimensions. *Nature Reviews. Clinical Oncology* 2013; **10**: 643–655.
12. Correa P and Houghton J. Carcinogenesis of *Helicobacter pylori*. *Gastroenterology* 2007; **133**: 659–672.
13. Helicobacter and Cancer Collaborative Group. Gastric cancer and *Helicobacter pylori*: a combined analysis of 12 case control studies nested within prospective cohorts. *Gut* 2001; **49**: 347–353.
14. Uemura N, Okamoto S, Yamamoto S, Matsumura N, Yamaguchi S, Yamakido M, *et al.* *Helicobacter pylori* infection and the development of gastric cancer. *The New England Journal of Medicine* 2001; **345**: 784–789.
15. Wroblewski LE and Peek RM Jr. *Helicobacter pylori* in gastric carcinogenesis: mechanisms. *Gastroenterology Clinics of North America* 2013; **42**: 285–298.
16. Hayakawa Y, Ariyama H, Stancikova J, Sakitani K, Asfaha S, Renz BW, *et al.* Mist1 expressing gastric stem cells maintain the normal and neoplastic gastric epithelium and are supported by a perivascular stem cell niche. *Cancer Cell* 2015; **28**: 800–814.
17. Carneiro F, Huntsman DG, Smyrk TC, Owen DA, Seruca R, Pharoah P, *et al.* Model of the early development of diffuse gastric cancer in E-cadherin mutation carriers and its implications for patient screening. *The Journal of Pathology* 2004; **203**: 681–687.
18. Yakirevich E and Resnick MB. Pathology of gastric cancer and its precursor lesions. *Gastroenterology Clinics of North America* 2013; **42**: 261–284.
19. Pharoah PD, Oliveira C, Machado JC, Keller G, Vogelsang H, Laux H, *et al.* CDH1 c-160a promotor polymorphism is not associated with risk of stomach cancer. *International Journal of Cancer*. 2002; **101**: 196–197.
20. Hansford S, Kaurah P, Li-Chang H, Woo M, Senz J, Pinheiro H, *et al.* Hereditary diffuse gastric cancer syndrome: CDH1 mutations and beyond. *Journal of American Medical Association Oncology* 2015; **1**: 23–32.
21. Fitzgerald RC, Hardwick R, Huntsman D, Carneiro F, Guilford P, Blair V, *et al.* Hereditary diffuse gastric cancer: updated consensus guidelines for clinical management and directions for future research. *Journal of Medical Genetics* 2010; **47**: 436–444.
22. Watson P, Vasen HF, Mecklin JP, Bernstein I, Aarnio M, Jarvinen HJ, *et al.* The risk of

- extra-colonic, extra-endometrial cancer in the Lynch syndrome. *International Journal of Cancer* 2008; **123**: 444–449.
23. Worthley DL, Phillips KD, Wayte N, Schrader KA, Healey S, Kaurah P, *et al.* Gastric adenocarcinoma and proximal polyposis of the stomach (GAPPS): a new autosomal dominant syndrome. *Gut* 2012; **61**: 774–779.
  24. Chun N and Ford JM. Genetic testing by cancer site: stomach. *Cancer Journal* 2012; **18**: 355–363.
  25. Scanziani E, Giusti AM, Gualtieri M and Fonda D. Gastric carcinoma in the Belgian shepherd dog. *Journal of Small Animal Practice* 1991; **32**: 465–469.
  26. Kurihara M, Shirakabe H, Izumi T, Miyasaka K, Yamaya F, Maruyama T, *et al.* Adenocarcinomas of the stomach induced in beagle dogs by oral administration of N-ethyl-N'-nitro-N-nitrosoguanidine. *Zeitschrift für Krebsforschung und klinische Onkologie* 1977; **90**: 241–252.
  27. Lubbes D, Mandigers PJ, Heuven HC and Teske E. Incidence of gastric carcinoma in Dutch Tervueren shepherd dogs born between 1991 and 2002. *Tijdschrift voor Diergeneeskunde* 2009; **134**: 606–610.
  28. Lecoindre P, Bystricka M, Chevallier M and Peyron C. Gastric carcinoma associated with Menetrier's-like disease in a West Highland white terrier. *The Journal of Small Animal Practice* 2012; **53**: 714–718.
  29. Munday JS, Aberdein D, Cullen GD and French AF. Menetrier disease and gastric adenocarcinoma in 3 Cairn terrier littermates. *Veterinary Pathology* 2012; **49**: 1028–1031.
  30. Kolbjornsen O, Press CM and Landsverk T. Gastropathies in the Lundehund. I. Gastritis and gastric neoplasia associated with intestinal lymphangiectasia. *Acta Pathologica, Microbiologica, et Immunologica Scandinavica* 1994; **102**: 647–661.
  31. Qvigstad G, Kolbjornsen O, Skancke E and Waldum HL. Gastric neuroendocrine carcinoma associated with atrophic gastritis in the norwegian lundehund. *Journal of Comparative Pathology* 2008; **139**: 194–201.
  32. Amorim I, Taulescu MA, Ferreira A, Rema A, Reis CA, Faustino AM, *et al.* An immunohistochemical study of canine spontaneous gastric polyps. *Diagnostic Pathology* 2014; **9**: 166. doi:10.1186/s13000-014-0166-z.
  33. Amorim I, Freitas DP, Magalhães A, Faria F, Lopes C, Faustino AM, *et al.* A comparison of *Helicobacter pylori* and non-*Helicobacter pylori* *Helicobacter* spp. Binding to canine gastric mucosa with defined gastric glyco phenotype. *Helicobacter* 2014; **19**: 249–259.
  34. Cattoli G, van Vugt R, Zanoni RG, Sanguinetti V, Chiocchetti R, Gualtieri M, *et al.* Occurrence and characterization of gastric *Helicobacter* spp. in naturally infected dogs. *Veterinary Microbiology* 1999; **70**: 239–250.
  35. Hermanns W, Kregel K, Breuer W and Lechner J. *Helicobacter*-like organisms: histopathological examination of gastric biopsies from dogs and cats. *Journal of Comparative Pathology* 1995; **112**: 307–318.
  36. Amorim I, Smet A, Alves O, Teixeira S, Saraiva AL, Taulescu M, *et al.* Presence and significance of *Helicobacter* spp. in the gastric mucosa of Portuguese dogs. *Gut Pathogens* 2015; **7**: 12. doi:10.1186/s13099-015-0057-1. eCollection 2015.
  37. Jemal A, Bray F, Center MM, Ferlay J, Ward E and Forman D. Global cancer statistics. *CA: A Cancer Journal for Clinicians* 2011; **61**: 69–90.
  38. Zanghieri G, Di Gregorio C, Sacchetti C, Fante R, Sassatelli R, Cannizzo G, *et al.* Familial occurrence of gastric cancer in the 2-year experience of a population-based registry. *Cancer* 1990; **66**: 2047–2051.
  39. McLean MH and El-Omar EM. Genetics of gastric cancer. *Nature Reviews Gastroenterology & Hepatology* 2014; **11**: 664–674.
  40. Sereno M, Aguayo C, Guillen Ponce C, Gomez-Raposo C, Zambrana F, Gomez-Lopez M, *et al.* Gastric tumours in hereditary cancer syndromes: clinical features, molecular biology and strategies for prevention. *Clinical & Translational Oncology* 2011; **13**: 599–610.
  41. Shah MA and Ajani JA. Gastric cancer – an enigmatic and heterogeneous disease. *Journal of American Medical Association* 2010; **303**: 1753–1754.
  42. Fenoglio-Preiser C, Muñoz N, Carneiro F, Powell SM, Correa P, Rugge M, *et al.* Tumours of the stomach. In: *World Health Organization Classification of Tumours. Pathology and Genetics of Tumours of the Digestive System*. SR Hamilton and LA Aaltonen Eds., Lyon, IARC Press, 2010: 38–52. <http://www.iarc.fr/en/publications/pdfs-online/pat-gen/bb2/>.
  43. Kelley JR and Duggan JM. Gastric cancer epidemiology and risk factors. *Journal of Clinical Epidemiology* 2003; **56**: 1–9.
  44. Bonenkamp JJ, van Krieken H, Craanen M and Van de Velde CJ. Gastric Cancer. In: *Oxford Textbook of Oncology*. 2nd edn., New York, Oxford University Press, 2002: 1517–1535.

45. Deng JY and Liang H. Clinical significance of lymph node metastasis in gastric cancer. *World Journal of Gastroenterology* 2014; **20**: 3967–3975.
46. Withrow SJ and Vail DM. Gastric cancer. In: *Small Animal Clinical Oncology*. 4th edn., Missouri, Saunders Elsevier, 2007: 480–483.
47. Patnaik AK, Hurvitz AI and Johnson GF. Canine gastric adenocarcinoma. *Veterinary Pathology* 1978; **15**: 600–607.
48. Willard MD. Alimentary neoplasia in geriatric dogs and cats. *The Veterinary Clinics of North America: Small Animal Practice* 2012; **42**: 693–706, vi.
49. Lamb CR and Grierson J. Ultrasonographic appearance of primary gastric neoplasia in 21 dogs. *The Journal of Small Animal Practice* 1999; **40**: 211–215.
50. von Babo V, Eberle N, Mischke R, Meyer-Lindenberg A, Hewicker-Trautwein M, Nolte I, *et al.* Canine non-hematopoietic gastric neoplasia. Epidemiologic and diagnostic characteristics in 38 dogs with post-surgical outcome of five cases. *Tierärztliche Praxis. Ausgabe K, Kleintiere/Heimtiere* 2012; **40**: 243–249.
51. Frost D, Lasota J and Miettinen M. Gastrointestinal stromal tumors and leiomyomas in the dog: a histopathologic, immunohistochemical, and molecular genetic study of 50 cases. *Veterinary Pathology* 2003; **40**: 42–54.
52. Sullivan M, Lee R, Fisher EW, Nash AS and McCandlish IA. A study of 31 cases of gastric carcinoma in dogs. *The Veterinary Record* 1987; **120**: 79–83.
53. Fonda D, Gualtieri M and Scanziani E. Gastric carcinoma in the dog: a clinicopathological study of 11 cases. *Journal of Small Animal Practice* 1989; **30**: 353–360.
54. Lingeman CH, Garner FM and Taylor DO. Spontaneous gastric adenocarcinomas of dogs: a review. *Journal of the National Cancer Institute* 1971; **47**: 137–153.
55. Seim-Wikse T, Kolbjornsen O, Jorundsson E, Benestad SL, Bjornvad CR, Grotmol T, *et al.* Tumour gastrin expression and serum gastrin concentrations in dogs with gastric carcinoma are poor diagnostic indicators. *Journal of Comparative Pathology* 2014; **151**: 207–211.
56. Carrasco V, Canfran S, Rodriguez-Franco F, Benito A, Sainz A and Rodriguez-Bertos A. Canine gastric carcinoma: immunohistochemical expression of cell cycle proteins (p53, p21, and p16) and heat shock proteins (Hsp27 and Hsp70). *Veterinary Pathology* 2011; **48**: 322–329.
57. Doster AR, Yhee JY, Kim JH, Im KS and Sur JH. CDX-2 and HER-3 expression in canine gastric and colorectal adenocarcinomas. *Journal of Comparative Pathology* 2011; **145**: 12–19.
58. Lee HC, Kim JH, Jee CH, Lee JH, Moon JH, Kim NH, *et al.* A case of gastric adenocarcinoma in a Shih Tzu dog: successful treatment of early gastric cancer. *The Journal of Veterinary Medical Science* 2014; **76**: 1033–1038.
59. Terragni R, Vignoli M, Rossi F, Laganga P, Leone VF, Graham JP, *et al.* Stomach wall evaluation using helical hydro-computed tomography. *Veterinary Radiology & Ultrasound* 2012; **53**: 402–405.
60. Lee JH, Kim JG, Jung HK, Kim JH, Jeong WK, Jeon TJ, *et al.* Clinical practice guidelines for gastric cancer in Korea: an evidence-based approach. *Journal of Gastric Cancer* 2014; **14**: 87–104.
61. Moehler M, Baltin CT, Ebert M, Fischbach W, Gockel I, Grenacher L, *et al.* International comparison of the German evidence-based S3-guidelines on the diagnosis and multimodal treatment of early and locally advanced gastric cancer, including adenocarcinoma of the lower esophagus. *Gastric Cancer* 2014; **18**: 550–563.
62. McLoughlin JM. Adenocarcinoma of the stomach: a review. *Proceedings (Baylor University. Medical Center)* 2004; **17**: 391–399.
63. Ezoe Y, Muto M, Uedo N, Doyama H, Yao K, Oda I, *et al.* Magnifying narrowband imaging is more accurate than conventional white-light imaging in diagnosis of gastric mucosal cancer. *Gastroenterology* 2011; **141**: 2017–2025. e3.
64. Serrano M, Kikuste I and Dinis-Ribeiro M. Advanced endoscopic imaging for gastric cancer assessment: new insights with new optics? *Best Practice & Research. Clinical Gastroenterology* 2014; **28**: 1079–1091.
65. Jackson C, Cunningham D, Oliveira J and ESMO Guidelines Working Group. Gastric cancer: ESMO clinical recommendations for diagnosis, treatment and follow-up. *Annals of Oncology* 2009; **20**(Suppl. 4): 34–36.
66. Waddell T, Verheij M, Allum W, Cunningham D, Cervantes A, Arnold D, *et al.* Gastric cancer: ESMO-ESSO-ESTRO clinical practice guidelines for diagnosis, treatment and follow-up. *European Journal of Surgical Oncology* 2014; **40**: 584–591.
67. Rivers BJ, Walter PA, Johnston GR, Feeney DA and Hardy RM. Canine gastric neoplasia: utility of ultrasonography in diagnosis. *Journal of the American Animal Hospital Association* 1997; **33**: 144–155.

68. Marolf AJ, Bachand AM, Sharber J and Twedt DC. Comparison of endoscopy and sonography findings in dogs and cats with histologically confirmed gastric neoplasia. *The Journal of Small Animal Practice* 2015; **56**: 339–344.
69. Terragni R, Vignoli M, van Bree HJ, Gaschen L and Saunders JH. Diagnostic imaging and endoscopic finding in dogs and cats with gastric tumors: a review. *Schweizer Archiv für Tierheilkunde* 2014; **156**: 569–576.
70. Eisele J, McClaran JK, Runge JJ, Holt DE, Culp WT, Liu S, *et al.* Evaluation of risk factors for morbidity and mortality after pylorotomy and gastroduodenostomy in dogs. *Veterinary Surgery* 2010; **39**: 261–267.
71. Sharman MJ, Bacci B, Whittam T and Mansfield CS. In vivo histologically equivalent evaluation of gastric mucosal topologic morphology in dogs by using confocal endomicroscopy. *Journal of Veterinary Internal Medicine* 2014; **28**: 799–808.
72. Lauren P. The two histological main types of gastric carcinoma: diffuse and so-called intestinal-type carcinoma. an attempt at a histo-clinical classification. *Acta Pathologica et Microbiologica Scandinavica* 1965; **64**: 31–49.
73. Hass HG, Smith U, Jager C, Schaffer M, Wellhauber U, Hehr T, *et al.* Signet ring cell carcinoma of the stomach is significantly associated with poor prognosis and diffuse gastric cancer (Lauren's): single-center experience of 160 cases. *Onkologie* 2011; **34**: 682–686.
74. Liu L, Wang ZW, Ji J, Zhang JN, Yan M, Zhang J, *et al.* A cohort study and meta-analysis between histopathological classification and prognosis of gastric carcinoma. *Anti-Cancer Agents in Medicinal Chemistry* 2013; **13**: 227–234.
75. Oliveira C, Seruca R and Carneiro F. Hereditary gastric cancer. *Best Practice & Research. Clinical Gastroenterology* 2009; **23**: 147–157.
76. de Vries AC, van Grieken NC, Looman CW, Casparie MK, de Vries E, Meijer GA, *et al.* Gastric cancer risk in patients with premalignant gastric lesions: a nationwide cohort study in the Netherlands. *Gastroenterology* 2008; **134**: 945–952.
77. Head KW. *Histological Classification of Tumors of the Alimentary System of Domestic Animals*. Published by the Armed Forces Institute of Pathology in cooperation with the American Registry of Pathology and the World Health Organization Collaborating Center for Worldwide Reference on Comparative Oncology, 2003.
78. Akiyama T, Sudo C, Ogawara H, Toyoshima K and Yamamoto T. The product of the human c-erbB-2 gene: a 185-kilodalton glycoprotein with tyrosine kinase activity. *Science* 1986; **232**: 1644–1646.
79. Zhang SY, Zhang SQ, Nagaraju GP and El-Rayes BF. Biomarkers for personalized medicine in GI cancers. *Molecular Aspects of Medicine* 2015; **45**: 14–27.
80. Ajani JA, Bentrem DJ, Besh S, D'Amico TA, Das P, Denlinger C, *et al.* Gastric cancer, version 2.2013: featured updates to the NCCN Guidelines. *Journal of the National Comprehensive Cancer Network* 2013; **11**: 531–546.
81. Bang YJ, Van Cutsem E, Feyereislova A, Chung HC, Shen L, Sawaki A, *et al.* Trastuzumab in combination with chemotherapy versus chemotherapy alone for treatment of HER2-positive advanced gastric or gastro-oesophageal junction cancer (ToGA): a phase 3, open-label, randomised controlled trial. *Lancet* 2010; **376**: 687–697.
82. Shen L, Shan YS, Hu HM, Price TJ, Sirohi B, Yeh KH, *et al.* Management of gastric cancer in Asia: resource-stratified guidelines. *The Lancet. Oncology* 2013; **14**: e535–e547.
83. Terragni R, Casadei Gardini A, Sabattini S, Bettini G, Amadori D, Talamonti C, *et al.* EGFR, HER-2 and KRAS in canine gastric epithelial tumors: a potential human model? *PLoS One* 2014; **9**: e85388.
84. Burrai GP, Tanca A, De Miglio MR, Abbondio M, Pisanu S, Polinas M, *et al.* Investigation of HER2 expression in canine mammary tumors by antibody-based, transcriptomic and mass spectrometry analysis: is the dog a suitable animal model for human breast cancer? *Tumour Biology* 2015; **36**: 9083–9091.
85. Kapitanovic S, Radosevic S, Slade N, Kapitanovic M, Anđelinovic S, Ferencic Z, *et al.* Expression of erbB-3 protein in colorectal adenocarcinoma: correlation with poor survival. *Journal of Cancer Research and Clinical Oncology* 2000; **126**: 205–211.
86. Li Q, Yuan Z and Cao B. The function of human epidermal growth factor receptor-3 and its role in tumors (Review). *Oncology Reports* 2013; **30**: 2563–2570.
87. Mazziotta RM, Borczuk AC, Powell CA and Mansukhani M. CDX2 immunostaining as a gastrointestinal marker: expression in lung carcinomas is a potential pitfall. *Applied Immunohistochemistry & Molecular Morphology* 2005; **13**: 55–60.
88. Moskaluk CA, Zhang H, Powell SM, Cerilli LA, Hampton GM and Frierson HF Jr. Cdx2 protein expression in normal and malignant human tissues: an immunohistochemical survey using

- tissue microarrays. *Modern Pathology* 2003; **16**: 913–919.
89. Lee KJ, Inoue M, Otani T, Iwasaki M, Sasazuki S, Tsugane S, *et al.* Gastric cancer screening and subsequent risk of gastric cancer: a large-scale population-based cohort study, with a 13-year follow-up in Japan. *International Journal of Cancer* 2006; **118**: 2315–2321.
  90. Leja M, You W, Camargo MC and Saito H. Implementation of gastric cancer screening – the global experience. *Best Practice & Research: Clinical Gastroenterology* 2014; **28**: 1093–1106.
  91. Jiang L, Yang KH, Chen Y, Guan QL, Zhao P, Tian JH, *et al.* Systematic review and meta-analysis of the effectiveness and safety of extended lymphadenectomy in patients with resectable gastric cancer. *The British Journal of Surgery* 2014; **101**: 595–604.
  92. Songun I, Putter H, Kranenbarg EM, Sasako M and van de Velde CJ. Surgical treatment of gastric cancer: 15-year follow-up results of the randomised nationwide Dutch D1D2 trial. *The Lancet.Oncology* 2010; **11**: 439–449.
  93. van der Post RS, Vogelaar IP, Carneiro F, Guilford P, Huntsman D, Hoogerbrugge N, *et al.* Hereditary diffuse gastric cancer: updated clinical guidelines with an emphasis on germline CDH1 mutation carriers. *Journal of Medical Genetics* 2015; **52**: 361–374.
  94. Allum WH, Blazeby JM, Griffin SM, Cunningham D, Jankowski JA, Wong R, *et al.* Guidelines for the management of oesophageal and gastric cancer. *Gut* 2011; **60**: 1449–1472.
  95. Sano T and Kodera Y. Japanese gastric cancer treatment guidelines 2010 (ver. 3). *Gastric Cancer* 2011; **14**: 113–123.
  96. Nielsen C and Anderson GM. Metastasis of gastric adenocarcinoma to the abdominal wall following placement of a gastrostomy tube in a dog. *The Canadian Veterinary Journal* 2005; **46**: 641–643.
  97. Sellon RK, Bissonnette K and Bunch SE. Long-term survival after total gastrectomy for gastric adenocarcinoma in a dog. *Journal of Veterinary Internal Medicine* 1996; **10**: 333–335.
  98. Janke L, Carlson CS and St Hill CA. The novel carbohydrate tumor antigen C2-O-sLe x is upregulated in canine gastric carcinomas. *Veterinary Pathology* 2010; **47**: 455–461.
  99. Gazvoda B, Juvan R, Zupanic-Pajnic I, Repse S, Ferlan-Marolt K, Balazic J, *et al.* Genetic changes in Slovenian patients with gastric adenocarcinoma evaluated in terms of microsatellite DNA. *European Journal of Gastroenterology & Hepatology* 2007; **19**: 1082–1089.
  100. Choi YY, Bae JM, An JY, Kwon IG, Cho I, Shin HB, *et al.* Is microsatellite instability a prognostic marker in gastric cancer? A systematic review with meta-analysis. *Journal of Surgical Oncology* 2014; **110**: 129–135.
  101. Zhu L, Li Z, Wang Y, Zhang C, Liu Y and Qu X. Microsatellite instability and survival in gastric cancer: a systematic review and meta-analysis. *Molecular and Clinical Oncology* 2015; **3**: 699–705.
  102. El-Omar EM. The importance of interleukin 1beta in Helicobacter pylori associated disease. *Gut* 2001; **48**: 743–747.
  103. He C, Tu H, Sun L, Xu Q, Gong Y, Jing J, *et al.* SNP interactions of Helicobacter pylori-related host genes PGC, PTPN11, IL1B, and TLR4 in susceptibility to gastric carcinogenesis. *Oncotarget* 2015; **6**: 19017–19026.
  104. Loh M, Koh KX, Yeo BH, Song CM, Chia KS, Zhu F, *et al.* Meta-analysis of genetic polymorphisms and gastric cancer risk: variability in associations according to race. *European Journal of Cancer* 2009; **45**: 2562–2568.
  105. Xue H, Lin B, Ni P, Xu H and Huang G. Interleukin-1B and interleukin-1 RN polymorphisms and gastric carcinoma risk: a meta-analysis. *Journal of Gastroenterology and Hepatology* 2010; **25**: 1604–1617.
  106. Persson C, Canedo P, Machado JC, El-Omar EM and Forman D. Polymorphisms in inflammatory response genes and their association with gastric cancer: a HuGE systematic review and meta-analyses. *American Journal of Epidemiology* 2011; **173**: 259–270.
  107. Qinghai Z, Yanying W, Yunfang C, Xukui Z and Xiaoqiao Z. Effect of interleukin-17A and interleukin-17 F gene polymorphisms on the risk of gastric cancer in a Chinese population. *Gene* 2014; **537**: 328–332.
  108. Rafiei A, Hosseini V, Janbabai G, Ghorbani A, Ajami A, Farzmandfar T, *et al.* Polymorphism in the interleukin-17A promoter contributes to gastric cancer. *World Journal of Gastroenterology* 2013; **19**: 5693–5699.
  109. Zheng L, Zhu C, Gu J, Xi P, Du J and Jin G. Functional polymorphism rs4072037 in MUC1 gene contributes to the susceptibility to gastric cancer: evidence from pooled 6,580 cases and 10,324 controls. *Molecular Biology Reports* 2013; **40**: 5791–5796.
  110. Sala N, Munoz X, Travier N, Agudo A, Duell EJ, Moreno V, *et al.* Prostate stem-cell antigen gene is associated with diffuse and intestinal gastric cancer in Caucasians: results from the EPIC-EURGAST

- study. *International Journal of Cancer* 2012; **130**: 2417–2427.
111. Study Group of Millennium Genome Project for Cancer, Sakamoto H, Yoshimura K, Saeki N, Katai H, Shimoda T, *et al.* Genetic variation in PSCA is associated with susceptibility to diffuse-type gastric cancer. *Nature Genetics* 2008; **40**: 730–740.
  112. Mocellin S, Verdi D, Pooley KA and Nitti D. Genetic variation and gastric cancer risk: a field synopsis and meta-analysis. *Gut* 2015; **64**: 1209–1219.
  113. Kakiuchi M, Nishizawa T, Ueda H, Gotoh K, Tanaka A, Hayashi A, *et al.* Recurrent gain-of-function mutations of RHOA in diffuse-type gastric carcinoma. *Nature Genetics* 2014; **46**: 583–587.
  114. Zang ZJ, Cutcutache I, Poon SL, Zhang SL, McPherson JR, Tao J, *et al.* Exome sequencing of gastric adenocarcinoma identifies recurrent somatic mutations in cell adhesion and chromatin remodeling genes. *Nature Genetics* 2012; **44**: 570–574.
  115. Oliveira C, Sousa S, Pinheiro H, Karam R, Bordeira-Carrico R, Senz J, *et al.* Quantification of epigenetic and genetic 2nd hits in CDH1 during hereditary diffuse gastric cancer syndrome progression. *Gastroenterology* 2009; **136**: 2137–2148.
  116. Guilford P, Hopkins J, Harraway J, McLeod M, McLeod N, Harawira P, *et al.* E-cadherin germline mutations in familial gastric cancer. *Nature* 1998; **392**: 402–405.
  117. Majewski IJ, Kluijt I, Cats A, Scerri TS, de Jong D, Kluin RJ, *et al.* An alpha-E-catenin (CTNNA1) mutation in hereditary diffuse gastric cancer. *The Journal of Pathology* 2013; **229**: 621–629.
  118. Schuetz JM, Leach S, Kaurah P, Jeyes J, Butterfield Y, Huntsman D, *et al.* Catenin family genes are not commonly mutated in hereditary diffuse gastric cancer. *Cancer Epidemiology, Biomarkers & Prevention* 2012; **21**: 2272–2274.
  119. Gaston D, Hansford S, Oliveira C, Nightingale M, Pinheiro H, Macgillivray C, *et al.* Germline mutations in MAP3K6 are associated with familial gastric cancer. *PLoS Genetics* 2014; **10**: e1004669.
  120. Wang J, Yang H, Shen Y, Wang S, Lin D, Ma L, *et al.* Direct sequencing is a reliable assay with good clinical applicability for KRAS mutation testing in colorectal cancer. *Cancer Biomarkers* 2013; **13**: 89–97.
  121. Lindblad-Toh K, Wade CM, Mikkelsen TS, Karlsson EK, Jaffe DB, Kamal M, *et al.* Genome sequence, comparative analysis and haplotype structure of the domestic dog. *Nature* 2005; **438**: 803–819.